

authentic material, cochromatography (TLC-3 solvents), IR and UV analysis. The results are tabulated below

TABLE 1 SURVEY OF FLAVONOIDS IN TEN *Liatris* SPECIES

Plant	Kaempferitrin	Flavonoids		Kaempferol	Quercetin
		Vicenin	Isoquercitrin		
<i>L. provincialis</i>	+	—	—	—	—
<i>L. pycnostachya</i>	+	—	+	+	—
<i>L. punctata</i>	+	—	—	+	—
<i>L. chapmanii</i> *	+	V-2	—	+	—
<i>L. secunda</i>	+	V-2	—	+	—
<i>L. graminifolia</i>	+	—	—	—	—
<i>L. gracilis</i>	—	V-2	Rutin	—	+
<i>L. spicata</i>	—	—	Rutin	—	—
<i>L. tenuifolia</i> *	+	V-1 + V-2	—	—	—
<i>L. elegans</i>	+	V-1 + V-2	—	—	—

\* These were selected as typical examples for isolation

As can be seen from the above table, kaempferitrin is present in all species except in *L. gracilis* and *L. spicata*. Except for rutin and some flavonoid-like substances which were present only in traces in *L. spicata*, none of the compounds mentioned by Kagan<sup>1</sup> could be seen in our sample. It is also worth mentioning here that the occurrence of kaempferitrin in 8 *Liatris* species is the second report of kaempferitrin in Compositae, the first being the observation that it occurs in *Notonia grandiflora*.<sup>7</sup> Vicenin-1 which has been found in 5 of the *Liatris* species is still not a commonly-occurring glycoside although it has been synthesized and its structure thoroughly established by Bouillant and Chopin.<sup>4</sup>

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<sup>7</sup> RAO, D. V. and RAO, E. V. (1972) *Planta Med.* **22**, 205.

## LONICEROSIDE (SECOLOGANIN) IN *CORNUS OFFICINALIS* AND *C. MAS*

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**Key Word Index**—*Cornus officinalis*, *C. mas*, Cornaceae, loniceroside (secologanin), iridoid glucosides.

**Plants** *Cornus officinalis* Sieb. et Zucc. and *C. mas* L. (sect. *Macrocarpum* Spach).<sup>1</sup>  
**Source** Hørsholm Arboretum, Denmark. **Previous work on iridoid glucosides** In *C.*

<sup>1</sup> WANGERIN, W. (1910) in *Das Pflanzenreich* (Engler, A., ed.), Vol. IV, p. 229, Engelmann, Leipzig.

*officinalis*: loganin and morroniside,<sup>2</sup> morroniside (in fruits)<sup>3</sup> *Present work* Acetylation of an iridoid glycoside fraction (700 mg), obtained by the method previously described<sup>4</sup> from frozen leaves (300 g, collected in September, processed in December, 1971) of *C. officinalis*, followed by preparative TLC separation (silica gel-Et<sub>2</sub>O-C<sub>6</sub>H<sub>6</sub>) of the reaction mixture, yielded, as the major product, the tetraacetate of loniceroside<sup>5</sup> (secologanin<sup>6</sup>), a glucoside previously encountered in leaves of *Lonicera morrowii* A. Gray (Caprifoliaceae)<sup>5</sup> The non-crystalline tetraacetate, exhibiting the expected and almost completely interpreted NMR spectrum, crystallized on seeding with an authentic specimen of loniceroside tetraacetate, kindly provided by Professor Mitsuhashi, Hokkaido University, Sapporo, Japan The purified product melted at 111–112°, alone or in admixture with the authentic specimen (reported<sup>5</sup> m p 115–116°) Similar processing of leaves of *C. mas* gave identical results

Feeding experiments previously established that loniceroside is a precursor for morroniside in fruits of *C. officinalis*<sup>3</sup> The present finding ascertains that loniceroside is, in fact, a true intermediate on the pathway from loganin to morroniside

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<sup>2</sup> ENDO, T. and TAGUCHI, H. (1970) *Lecture*, quoted after Ref. 3

<sup>3</sup> INOUE, H., UEDA, S. and TAKEDA, Y. (1971) *Tetrahedron Letters* 4069

<sup>4</sup> ROSENDAL JENSEN, S., KJÆR, A. and JUHL NIELSEN, B. (1973) *Acta Chem. Scand.* 27, in press

<sup>5</sup> SOUZU, I. and MITSUHASHI, H. (1970) *Tetrahedron Letters* 191

<sup>6</sup> BATTERSBY, A. R., BURNETT, A. R. and PARSONS, P. G. (1968) *Chem. Commun.* 1280

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## GENIPOSIDE AND MONOTROPEIN IN *CORNUS SUECICA*

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**Key Word Index**—*Cornus suecica*, Cornaceae, geniposide, monotropein, iridoid glucosides

*Plant* *Cornus suecica* L. (subgenus *Arctocrania* Endl.)<sup>1</sup> *Source* Rold Skov, Denmark *Previous work* Aucubin, weak reaction on paper chromatography<sup>2</sup> *Present work*. Whole frozen plants (385 g) were extracted with 75% EtOH. The water-soluble part was extracted with BuOH (4 × 30 ml) and divided into a soluble fraction, *A* (4.1 g), and a residue, *B* (14.0 g). After treatment with Al<sub>2</sub>O<sub>3</sub>,<sup>3</sup> *A* gave 0.38 g of mixture, purified by preparative TLC (CHCl<sub>3</sub>-MeOH, 3:1). Two recrystallizations from wet EtOAc of the major fraction (144 mg) afforded pure geniposide (46 mg), m p 161–162°, [α]<sub>D</sub><sup>23</sup> +8.7° (c 2.2, H<sub>2</sub>O) [lit. values<sup>4</sup> m p 163–164°, [α]<sub>D</sub> +7.5°, H<sub>2</sub>O], identified by its characteristic <sup>1</sup>H NMR

<sup>1</sup> WANGERIN, W. (1910) in *Das Pflanzenreich* (Engler, A., ed.), Vol. IV, p. 1, Engelmann, Leipzig

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<sup>4</sup> INOUE, H., SAITO, S., TAGUCHI, H. and ENDO, T. (1969) *Tetrahedron Letters* 2347